

# Metadata of the chapter that will be visualized in SpringerLink

Book Title	Transforming Coastal Zone for Sustainable Food and Income Security	
Series Title		
Chapter Title	Development of Ionome (Salt-Omic) for the Varietal Improvement and Food Security of the Coastal Region of India	
Copyright Year	2022	
Copyright HolderName	The Author(s), under exclusive license to Springer Nature Switzerland AG	
Corresponding Author	Family Name	<b>Mondal</b>
	Particle	
	Given Name	<b>Bidisha</b>
	Prefix	
	Suffix	
	Role	
	Division	
	Organization	The Neotia University
	Address	743368, Sarisha, West Bengal, India
	Email	bidisha.mondal@tnu.in
Abstract	<p>Global agricultural productivity is regulated by soil salinity, one of the major abiotic constraints faced by the farmers, growers and breeders. The genomic, transcriptomics, proteomics and metabolomic salt profile of coastal plants could provide an insight into the mechanisms by which the differential performance is regulated in contrasting varieties of a single crop. This study proposes the construction of an Ionome suitable for the coastal saline region for sustainable food security. This study focuses on functional genomic studies of saline belt crops and meta-analysis of the information for proposed Ionome (Salt-Omic) development. In the salt-genome segment, the appropriate genes were identified and categorized covering ion-transport-genes, senescence-associated genes, molecular-chaperones, dehydration-genes. Proteome provides additional information on protein coding sequences, endogenous small molecules. The identified genes, proteins and signalling pathways could form an Ionome repository for molecular crop breeding programmes. The primary bioinformatics web source along with a customized database for several crops were found useful for identifying essential biomolecules. The study was able to assist in the formation of agri-ionome for the improvement of coastal crops. The sequential integration of agri-engineering model along with Omic details could be utilized for the construction of an explicit repository for molecular plant breeders in a way similar to AMBAB, LIS, Pulsechip or RiceMetaSys.</p>	
Keyword (separated by '-')	Ionome (Salt-omic) - Molecular crop-improvement - Food security - Breeder focused-database	

# Chapter 5

## Development of Ionome (Salt-Omic) for the Varietal Improvement and Food Security of the Coastal Region of India



**Bidisha Mondal**

**Abstract** Global agricultural productivity is regulated by soil salinity, one of the major abiotic constraints faced by the farmers, growers and breeders. The genomic, transcriptomics, proteomics and metabolomic salt profile of coastal plants could provide an insight into the mechanisms by which the differential performance is regulated in contrasting varieties of a single crop. This study proposes the construction of an Ionome suitable for the coastal saline region for sustainable food security. This study focuses on functional genomic studies of saline belt crops and meta-analysis of the information for proposed Ionome (Salt-Omic) development. In the salt-genome segment, the appropriate genes were identified and categorized covering ion-transport-genes, senescence-associated genes, molecular-chaperones, dehydration-genes. Proteome provides additional information on protein coding sequences, endogenous small molecules. The identified genes, proteins and signalling pathways could form an Ionome repository for molecular crop breeding programmes. The primary bioinformatics web source along with a customized database for several crops were found useful for identifying essential biomolecules. The study was able to assist in the formation of agri-ionome for the improvement of coastal crops. The sequential integration of agri-engineering model along with Omic details could be utilized for the construction of an explicit repository for molecular plant breeders in a way similar to AMBAB, LIS, Pulsechip or RiceMetaSys.

**Keyword** Ionome (Salt-omic) · Molecular crop-improvement · Food security · Breeder focused-database

### 5.1 Introduction

Salinity problem creates massive impact on food security of our country. Enhanced crop productivity and area augmentation are the two focus area that could assist in sustainable food security. India requires 311 million food grains for its exceedingly

---

B. Mondal (✉)  
The Neotia University, 743368, Sarisha, West Bengal, India  
e-mail: [bidisha.mondal@tnu.in](mailto:bidisha.mondal@tnu.in)

© The Author(s), under exclusive license to Springer Nature Switzerland AG 2022  
T. D. Lama et al. (eds.), *Transforming Coastal Zone for Sustainable Food and Income Security*, [https://doi.org/10.1007/978-3-030-95618-9\\_5](https://doi.org/10.1007/978-3-030-95618-9_5)

1

growing population of approximately 1.43 billion in 2030. Intensive cropping, biodiversity loss, abundant fertilizer application, ground water depletion, deforestation, increased urbanization has led to an extraordinary increase in soil salinization in more than 6.74 million ha of land in the country. Advanced soil reclamation using novel technologies and strategies recovered and increased the crop area transforming the affected 2.18 million hectares of salt affected soil (Kumar and Sharma 2020). Inclusive technologies of gypsum-based treatment of sodic soil, subsurface drainage of saline soil, phytoremediation, bio-remediation, agro-forestry, novel breeding techniques could increase food security, income elevation and rural employability. The selection, retention and promotion of advanced breeding technologies could boost crop productivity aiding food security. The economically important varieties of rice, sunflower, sesame, moong, mustard, jute, vegetables and betel leaf grown in the coastal saline belt of West Bengal, India are susceptible to salinity stress. Plants can tolerate salinity stress by processes including salt exclusion or inclusion. The most predominant form of salt stress involving NaCl causes nutrient and metabolic imbalance. Increased salt stress leads to ion toxicity affecting the water retention capacity of the plant.

The breeders face constant challenges in development of new varieties resilient to extreme abiotic stresses including salinity. In such a condition unique breeding techniques will focus on expanding the gene pool or introspection of the present gene pool to identify some potential well performers. In recent times a comprehensive bioinformatics study involving inclusive approach covering all omic information could be applied for developing dynamic breeding strategies. The inclusion of high throughput techniques of DNA/RNA sequencing, microarray data and advanced proteomics could help the breeders to enter inside the unexplored territories of molecular crop advancement programme.

In salt and salinity management a number of agri-engineering computational models were already been invented and used. Scientists have analysed water flow and salt flow processes and developed scientific and engineering models for salinity management. On the basis of scope of application two models, basin model and field-scale models were developed by soil scientists, agri-engineers and agri-informatic professionals. LEACHC, SWAP, SOWACH, HYDRUS and UNSATCHEM were largely used for analysis of water flow, salt flow and plant performance. Among these models SWAP software package involves one-dimensional vertical transport of water, solute and heat and could predict crop yield. The SWAP model was effective in irrigation application and effective yield return studies in cotton (Pan et al. 2020).

SOWACH, a third generation salinity management model was written using QBASIC and runs under DOS and WINDOWS operating system. In a test, it was proved SOWACH model could effectively identify the rooting architecture ideal for increased yield (Alahmad et al. 2019). This model in alfalfa proved that across salinity treatments, final root length density was 24% higher in high fibrous root type and accordingly herbage yield of these root type superseded the low fibrous type by 14%.

The UNSATCHEM model was able to evaluate the efficacy of gypsum administered at different depths and able to suggest the feasibility of alternative management practises. Green manuring involving calcite could reclaim sodic field with less

71 water or without addition of gypsum was proved by software application (Amini  
72 et al. 2016). The abundant application of agri-engineering computer software for  
73 analysing saline soil was invented for saving time, expense and accuracy of the  
74 growers, farmers and stakeholders.

75 The apparent abundance of soil engineering models could be surpassed if infor-  
76 mation on molecular crop databases could be utilized directly in salinity research.  
77 In reality, the employment of biological database in salinity research is meagre. The  
78 vast molecular resource available in public domain could be effectively used for  
79 Ionome construction and allied research. In this aspect the application and perfor-  
80 mance of OMIC data have enormous potential. The available OMIC database related  
81 to salinity could be exploited by plant breeders and molecular biologists. The key  
82 information for plant Ionome construction could be gathered from three main public  
83 repositories. GenBank, EMBL and DDBJ along with UniProt and Swissprot could  
84 provide initial data for salinity gene and protein sequence information. The major  
85 sequence repositories includes curated collections and provide holistic information  
86 on different domains (Lizumi and Sakai 2020). The individuality of these collections  
87 are maintained by their virtue of curation as well as unique approach of presentation  
88 of biological discoveries. A General plant database search could provide the detailed  
89 idea about available plant-omic information. Additionally, sequential exploration  
90 of the customized resources could assist in plant Ionome construction. The identi-  
91 fication of important salinity gene, protein, EST, biological pathways, conserved  
92 domains and other relevant information were discussed in this paper to emphasize  
93 the importance of crop Ionome construction. Ionome (Salt-OMIC) could assist in the  
94 speedy selection and propagation of salt tolerant and resistant crops for the rejuvena-  
95 tion of the coastal and inland saline belts of developing countries ultimately ensuring  
96 food security for the human population.

## 97 5.2 Materials and Methods

98 In the present study two approaches were taken for OMIC data collection. In the first  
99 approach all the available primary bioinformatics resource were explored for devel-  
100 opment of proposed Ionome relevant to crop improvement programme of saline belt  
101 of West Bengal, India. In this study mainly GenBank, DDBJ and EMBL, Uniprot and  
102 Swissprot repositories were searched for gathering OMIC information of the major  
103 crops grown in saline belt of West Bengal. Additionally several other customised  
104 public database for the above mentioned crops were investigated for gene and cDNA.

105 In second approach the open source research database Google Scholar was used  
106 for article searching using a search string of salt + omic + crop name consensus  
107 tab with only one variable (crop name). It yielded articles with a sample size of  
108 9818. Only English articles published in last 5 years were included. The selected  
109 articles were screened on the basis of abstract and keywords. In initial selection grey  
110 literature, preprints, presentation, duplication and keynotes were excluded manually  
111 reducing the sample size to 303. In the next phase the study applied Voyant tool

**Table 5.1** Selection and screening of salt-omic-based research articles

Crop	Extracted paper (n)	Initial selection	Selected full paper
Rice	5010	254	14
Lentil	1240	57	9
Mung	1480	62	9
Sunflower	748	74	11
Mustard	514	102	9
Sesame	384	37	10
Jute	462	117	8
Total	9818	703	70

112 (Sampsel 2018), an open source, web-based application for executing text mining  
 113 and analysis, promoting rapid analysis of scholarly articles and interpretation of  
 114 texts from the selected articles. It precisely extracted the frequencies of keywords  
 115 from all selected published articles and ultimately 70 articles covering only salt-omic  
 116 information were thoroughly studied and data was extracted (Table 5.1).

## 117 5.3 Results and Discussion

### 118 5.3.1 *Exploitation of Computational Data Source for Ionome* 119 *Construction*

120 In the saline coastal belts of West Bengal apart from conventional rice cultivation  
 121 moong, mustard, sunflower, sesame, betel leaf and jute were also cultivated. In this  
 122 study, the unique dichotomous approach recorded major salt genes and proteins. A  
 123 comparative study was made between the crop specific saline data source and omic  
 124 associated papers. This study generated significant findings worthy to be applicable  
 125 for Ionome construction for coastal saline agricultural crops of West Bengal as well  
 126 as other salt affected regions of India. Omic information was extracted from reported  
 127 genes and proteins to be used as candidate or reference gene in abiotic stress tolerance.  
 128 In rice a Google Scholar a search using Salt + omic + rice shows 5010 papers with 22  
 129 salt genes reported in gene repositories. Collectively, 1480 omic papers were found  
 130 related to moong bean though there is no reporting of any gene. In NCBI platform a  
 131 database search for moong bean yields only 1 database recovery under nucleotides 2  
 132 reporting was found and that also comes under a test result performed for exclusive  
 133 NCBI internal data testing. Thousand two hundred forty papers were found for lentil  
 134 salt genes with 5 proteins. Whereas a search with salinity genes in soybean extracts  
 135 information from 10 databases out of which 8 could yield good result in reference  
 136 OMIC study of moong bean and legume breeding. In sunflower 1480 omic papers are  
 137 available with only one salt gene. NCBI covers a total of 22 bioinformatics database

138 under which 10 were selected most suitable for the collection of Ionome information.  
139 These information could be used by plant breeders for novel molecular studies with  
140 allied species or varieties and in integration could act as a valid resource for saline  
141 agricultural research. Crop specific useful salinity details under each database was  
142 recorded for salt breeding experiments. The database information of 10 cultivated  
143 saline belt crops were given in Table 5.2.

144 As rice is an important cereal the omic information were explored from several  
145 fields of publically accessible databases like OryzaExpress (Kudo et al. 2017), Oryz-  
146 abase (Kurata et al. 2006) and RiceXPro (Sato et al. 2013). The above domains were  
147 consulted for Ionome study. Salinity related 22 genes were found to be reported  
148 in NCBI database. The reported genes mainly came from japonica subspecies of  
149 rice and the available data could be utilized for advance screening of promising rice  
150 genotypes for a new region before the commencement of field trials. In RiceMetaSys  
151 web interface differential gene expression studies integrated molecular data for rice  
152 abiotic and biotic stresses (Sandhu et al. 2017). The breeders could use the Oryza-  
153 Express or Oryzabase for the construction of primers and contribute towards rice  
154 omic study as well. These database information could be utilized by the breeders of  
155 any omic discipline in the elucidation of molecular experimental result in rice. These  
156 database information could be utilized for pan-genomic and comparative candidate  
157 salt related genetic performance analysis of different salt tolerant indigenous and  
158 hybrid rice varieties.

159 LIS or legume information system is a community resource utilized for legume  
160 improvement programme (Gonzales et al. 2005). The LIS contains legume mines  
161 integrating 8 individual mines covering bean mine, chickpea mine, cowpea mine,  
162 medic mine, lupin mine, soy mine, peanut mine and joint vetch mines combining  
163 genomic and expression data of the inter-mines. LIS has potential to be used for gene  
164 identification along with the primary data available in major public database. Pulsedb,  
165 a database mapping software (Humann et al. 2019) and KnowPulse a diversity based  
166 software (Sanderson et al. 2019) could be utilized for stress related classical and  
167 molecular genetic studies of pulse.

168 BASC is an integrated bioinformatics system for Brassica research is available for  
169 omic study of salinity tolerant and resistant brassica (Timothy et al. 2007). BASC is  
170 generated for browsing and mining of Brassica genetic, genomic and phenotypic data.  
171 Multinational Brassica Genome Sequencing Project is a collection of five distinct  
172 modules, ESTDB, Microarray, MarkerQTL, CMap and EnSEMBL. ESTDB is a  
173 microarray module hosting expressed gene sequences and related annotation derived  
174 from comparison with GenBank, UniRef and the genome sequence of Arabidopsis.  
175 In mustard 130 salinity genes were evidenced with 720 proteins and 21EST from  
176 NCBI database. The Brassica database contains 47,555 unigenes made up of 17,939  
177 consensus and 29,616 singleton sequences requires separation of stress genes of  
178 Brassica. Additionally, raw and normalized both types of data could be extracted  
179 from BASC.

180 In tomato different database extracts noteworthy information for genetic expres-  
181 sion studies of tomato. Newly developed curated, open source integrated data  
182 resource including TGRD-Tomato Genomic Resources Database (Suresh et al.

**Table 5.2** Omic details of essential salinity database of major crops of saline coastal zone of West Bengal obtained from NCBI public database (NCBI 2016)

Crop	Gene	Nucleotide	Protein	EST	Bio-projects	Geo dataset	Popset	dbGap	Conserved domain	SRA
Rice	22	358	224	203	49	80	4	0	2	4
Sunflower	0	8	5	18	1	1	0	0	0	0
Moong	1	4	10	0	1	0	0	0	0	0
Lentil	0	5	5	0	1	2	0	3	1	1
Betel leaf	0	0	10	0	0	0	0	0	0	0
Mustard	130	85	720	21	1	13	0	13	0	5
Grass pea	2	462	329	0	2	529	4	0	0	12
Sesame	36	14	268	1	2	29	0	5	0	6
Jute	0	1	168	2	2	0	0	0	0	1
Soybean	11	53	30	26	8	9	1	2	1	0

183 2014), TFGD-Tomato Functional Genomics Database (Tranchida-Lombardo et al.  
184 2018), KaTomicsDB-Kazuka Tomato Genomics Database (Shirasawa et al. 2013),  
185 MoToDB-Metabolome Database (Grenan 2009) and COXPathDB (Narise et al.  
186 2017) were of prime importance for tomato omic studies. Gramene (Tello-Ruiz et al.  
187 2021) and MaizeGDB (Lawrence et al. 2008) portal was found equally effective in  
188 comparative functional genomic studies. The portal initially provided information for  
189 grass species but later the expansion of the portal made space for dicot species also.  
190 This portal hosts omic information of 93 reference genome of different plant species  
191 and 3.9 million genes covering 122,947 families with orthologous and paralogous  
192 details (Carson et al. 2016).

### 193 5.3.2 *Salt Gene and Protein for Ionome Construction*

194 In this study integrative meta-analysis efficiently identified several promising salinity  
195 genes. Twenty-five productive salt genes were identified in this study that could act  
196 as reference gene in virtual molecular breeding design. Bioinformatics tools could  
197 aid the breeder in initial design of a new crop related study revealing paralogous or  
198 ortholog gene function. The information presented in Tables 5.3 and 5.4 could be  
199 utilized for an inclusive study of sequence information, marker assay, trait associa-  
200 tion, locus details and biochemical path analysis of salinity tolerant important crops  
201 of West Bengal. This kind of bioinformatics data was utilized in testing of salinity  
202 responsive candidate gene in tomato for varietal performance testing (J'afar et al.  
203 2018). In another paper two drought tolerant landraces were compared using 122  
204 candidate gene studies. The approach identified high effect SNPs, structural variants  
205 and promising heat shock proteins and Cation /H<sup>+</sup> antiporters. Additionally, pan-  
206 genomic studies using 753 accessions in tomato showed 4873 genes were diverse  
207 from the reference genome (Chaudhary et al. 2019). In case of betel leaf the recorded  
208 information in NCBI repository requires huge input and there is provision for omic  
209 study.

### 210 5.3.3 *Signalling Pathways for Ionome Construction*

211 To combat salt stress the plant maintains an equilibrium among carbon sink distribu-  
212 tion, energy allocation and osmotic balance. Multiple signalling pathways become  
213 active for controlling salinity stress (Othman et al. 2017). Ionic stress signalling  
214 pathways along with osmotic stress signalling and detoxification pathways become  
215 operative in salt-stressed plants. A plethora of genes and proteins are related to  
216 this signalling cascades. SOS response, jasmonate pathway, tyrosine signalling,  
217 aquaporin regulation, ethylene synthesis, annexin mediated conductance, expres-  
218 sion of asparagine synthetase, hydroxyl proline-rich glycoproteins, ROS and ABA  
219 related proteins expressions were vital in salt stress management. Along with the



**Table 5.3** Essential genes identified for reference salt genomic studies

Gene	Crop	Gene ID	Function	References
SUV3	<i>Oryza sativa</i> Ssp. Japonica	4,334,089	Delayed senescence, ATP-dependent RNA helicase	Macovei et al. (2017)
KAT (4386)	Sunflower, rice, maize, potato	110,864,580	Cytosolic cation homeostasis	DeLeon et al. (2015)
HKT1	Japonica rice	4,341,971	Regulation of Na toxicity under salt stress	Oda et al. (2018)
NHX	Barley	103,934,031	Na/H exchanger, Vacuolar transporter	Liangbo et al. (2018)
SOS family	Rice	4,341,015	SOS response peptidase	Rice Consortium (2003), Platten et al. (2013)
TIFY gene family	Brassicaceae	4,342,421	Jasmonate ZIM domain specific protein	Reddy et al. (2017)
CRY1b	Rice, Brassica	543,688 2,829,419	Cryptochrome 1b Melatonin biosynthesis	Hwang and Back (2021)
CDPK	Rice/Arabidopsis	819,282	Receptor kinase Ca/CaM	Reddy et al. (2017)
MAPK	Dicot/Monocot	541,618 (Maize)	Serine/threonine protein kinase, salt related oxidative stress	Kong et al. (2019)
GH3	Rice, Gossypium, Vitis	107,896,815 Gossypium	Lipid transporter protein, Tyrosine kinase signalling	Wong et al. (2019)
ERF4 OsSIRP1	Monocot, spinach, Arabidopsis	101,290,597 Triticum	Jasmonate, ethylene pathway	Zhang et al. (2020)
NAC	Wheat, rice, sorghum, sugarcane, mung	818,902 Arabidopsis	AtHB13 and JUB1 transcription regulator	Ebrahimian-Motlagh et al. (2018)
OBF1 (2021)	Maize, rice, Dendrobium	542,394 Maize	Zn finger ring domain, ocs element transcription factor	Alexandrov et al. (2009)

(continued)

**Table 5.3** (continued)

Gene	Crop	Gene ID	Function	References
JcAP2/ERFs 2381	Rice, physic nut, Arabidopsis	844,348	Leucine rich repeat family, stomatal development	Tang et al. (2016)
OsFd1/Saltol	Red rice	3,974,662	Ferredoxin 1 ETS, Na/K homeostasis	He et al. 2020
RSS3	<i>Oryza sativa</i> Ssp. Japonica	4,350,435	Jasmonate induced gene expression	To et al. (2019), Toda et al. (2013)
OsGTv2	<i>Oryza sativa</i> Ssp. Japonica	4,330,612	Mitochondrial aldehyde dehydrogenase, Salinity adaptation	Xie et al. (2020)
AFP1,ninja family protein	Sunflower, tomato, rapeseed	110,920,831	putative ethylene responsive binding factor-associated repression, Ninja family	Badouin et al. (2017)
TaHAG1	Bread wheat. Barley, Arabidopsis	119,339,917	Modulating ROS production and salinity regulation	Zheng et al. (2021)
OsLEA	<i>Oryza sativa</i> Ssp. Japonica	4,339,745	ABA induced antioxidant stresses	Rice Consortium (2003)
<i>OsPYL/RCAR7</i>	<i>Oryza sativa</i> Ssp. Japonica	2,829,419	Glutaredoxin, ABA receptor	Bhatnagar et al. (2020)
TaOFP family	Triticum	3,760,030	Tillering, water stress	Wang et al. (2020)
ZmPTPN	Maize, Arabidopsis	AT5G50670	ABA signalling AsA biosynthesis	Zhang et al. (2020)
AQP (cDNA)	Chickpea, lathyrus, lentil	DY475124	Aquaporin regulation	Mantri et al. (2007)
ASNS (cDNA)	Chickpea, gram, lentil, grass pea	DY475477	Asparagine synthetase (glutamine hydrolysing)	Mantri et al. (2007)

**Table 5.4** Essential proteins identified for reference proteomic studies from Uniprot repository

Protein	Accession	Length (AA)	Pathway	Crop	References
WRKY transcription factor WRKY71	Q6QHD1	348	Gibberellin signalling pathway	Rice	Zhang et al. (2020)
Jasmonate ZIM domain-containing protein 9	Q8GSI0	179	Jasmonic acid signalling pathway	Rice	Young et al. (2018)
SNF1-related protein kinase catalytic subunit alpha	Q38997	521	Serine/threonine protein kinase	Arabidopsis	Simon et al. (2018)
Bidirectional sugar transporter SWEET15	Q9FY94	292	Transmembrane transporter	Maize	Doidy et al. (2019)
Ascorbate oxidase	M4DUF2	570	Oxidoreductase, defense pathway	Rapeseed	Nudrat et al. (2017)
Mitogen activated protein kinase	ACJ31803	586	ABA signal transduction	Groundnut	Wang et al. (2016)
XERICO	KAG1363776	166	E3 ubiquitin-protein ligase	Coconut	Brugiere et al. (2017)
5MYB	ABI74688	348	Transcriptase	Cabbage	Wang et al. (2015)
JUNGBRUNNEN	Q 9SK55	375	TF, Central longevity regulator	Thalecress	Dudhate et al. (2021)
ABRC5	AAR06258	141	Ethylene responsive binding factor	Sunflower	Najafi et al. (2018)
RCAR3	5GWO_D	175	ABA receptor	Rice	Hyunmi et al. (2012)
DELLA	NP_001240948	523	Ethylene regulator	Tomato	Shohat et al. (2020)
SnRK2	AID23890	354	sucrose non-fermenting 1-related protein kinase 2	Cotton	Liu et al. (2017)
ANAC017	EFH67341	547	Transcription regulator	Thale cress	Meng et al. (2019)
AKT	AT3G49850	693	AKT kinase telomeric DNA binding protein		Xu et al. (2020)

220 salt genes several important salt proteins were found impressive for comparative  
221 proteomic studies. In proteomics 2D gel electrophoresis, MALDI-TOF, mass spec-  
222 troscopy (Moco et al. 2006) and western blot have proved effective in analysing stress  
223 response (Shinozaki et al. 2018). Differential response of 49 proteins was noticed  
224 under aluminium stress in tomato seedlings and 40 for silicon stress in tomato (Zhou  
225 et al. 2009; Muneer et al. 2015).

## 226 5.4 Conclusions

227 The web-based primary bioinformatics database provides abundant omic informa-  
228 tion for several crops assisting scientists and researchers in the construction of  
229 crop specific public or private databases. Salinity stress is regarded as one of the  
230 robust abiotic problems reducing global food productivity. The proper alleviation  
231 of salt stress requires an understanding of the activity or expression of mineral and  
232 trace elements forming an integrated database including agri-engineering-based plant  
233 modelling and multivariate omic information to provide a solution to farmers. The  
234 omic study could provide true insight into the physiological activity of plant organs,  
235 differential expression of genes and epigenetic regulations. The above study proposes  
236 the necessity of the formation of a breeder-focused standalone Ionome (Salt-OMIC)  
237 database or crop specific customised construction for ensuring global food security  
238 under ever changing environmental complications.

## 239 References

- 240 Alahmad S, El Hassouni K, Bassi FM et al (2019) A major root architecture QTL responding to  
241 water limitation in durum wheat. *Front Plant Sci* 10:436. <https://doi.org/10.3389/fpls.2019.00436>
- 242 Alexandrov NN, Brover VV, Freidin S et al (2009) Insights into corn genes derived from large-scale  
243 cDNA sequencing. *Plant Mol Bio* 169(1–2):179–94. <https://doi.org/10.1007/s11103-008-9415-4>
- 244 Amini S, Ghadiri H, Chen C et al (2016) Salt-affected soils, reclamation, carbon dynamics, and  
245 biochar: a review. *J Soils Sediments* 16:939–953. <https://doi.org/10.1007/s11368-015-1293-1>
- 246 Badouin H et al (2017) The sunflower genome provides insights into oil metabolism, flowering and  
247 Asterid evolution. *Nature* 546(7656):148–152. <https://doi.org/10.1038/nature22380>
- 248 Bhatnagar N, Kim R, Han S et al (2020) Ectopic expression of OsPYL/RCAR7, an ABA receptor  
249 having low signaling activity, improves drought tolerance without growth defects in rice. *Int J*  
250 *Mol Sci* 21(11):4163. <https://doi.org/10.3390/ijms21114163>
- 251 Brugière N, Zhang W, Xu Q et al (2017) Overexpression of RING domain E3 ligase ZmXerico1  
252 confers drought tolerance through regulation of ABA homeostasis. *Plant Physiol* 175(3):1350–  
253 1369. <https://doi.org/10.1104/pp.17.01072>
- 254 Carson MA, Ethalinda KC, John LP et al (2016) MaizeGDB update: new tools, data and interface  
255 for the maize model organism database. *Nucleic Acids Res* 44(D1):D1195–D1201. <https://doi.org/10.1093/nar/gkv1007>
- 256 Chaudhary J, Khatri P, Singla P et al (2019) Advances in omics approaches for abiotic stress  
258 tolerance in tomato. *Biology (basel)* 8(4):90. <https://doi.org/10.3390/biology8040090>

- 259 De Leon TB, Linscombe S, Gregorio G et al (2015) Genetic variation in Southern USA rice geno-  
 260 types for seedling salinity tolerance. *Front Plant Sci* 6:374. <https://doi.org/10.3389/fpls.2015.00374>
- 262 Doidy J, Vidal U, Lemoine R (2019) Sugar transporters in Fabaceae, featuring SUT MST and  
 263 SWEET families of the model plant *Medicago truncatula* and the agricultural crop *Pisum sativum*.  
 264 *Plos One* 14(9). <https://doi.org/10.1371/journal.pone.0223173>
- 265 Dudhate A, Shinde H, Yu P et al (2021) Comprehensive analysis of NAC transcription factor  
 266 family uncovers drought and salinity stress response in pearl millet (*Pennisetum glaucum*). *BMC*  
 267 *Genomics* 22:70. <https://doi.org/10.1186/s12864-021-07382-y>
- 268 Ebrahimian-Motlagh S, Ribone PA, Thirumalaikumar VP et al (2018) JUNGBRUNNEN1 Confers  
 269 drought tolerance downstream of the HD-Zip I transcription factor AtHB13. *Front Plant Sci*  
 270 8:2118. <https://doi.org/10.3389/fpls.2017.02118>
- 271 Gonzales MD, Archuleta E, Farmer A et al (2005) The legume information system (LIS): an  
 272 integrated information resource for comparative legume biology. *Nucleic Acids Res* 33(1):660–  
 273 665. <https://doi.org/10.1093/nar/gki128>
- 274 Grenan AK (2009) Mo To DB A metabolic database of Tomato. *Plant Physiol* 151(4):1701–1702.  
 275 <https://doi.org/10.1104/pp.109.900308>
- 276 Othman HC, Millar H, Taylor NL (2017) Connecting salt stress signalling pathways with salinity-  
 277 induced changes in mitochondrial metabolic processes in C3 plants. *Plant Cell Environ* 40:2875–  
 278 2905. <https://doi.org/10.1111/pce.13034>
- 279 He L, Li M, Qiu Z et al (2020) Primary leaf-type ferredoxin 1 participates in photosynthetic electron  
 280 transport and carbon assimilation in rice. *Plant J* 104(1):44–58. <https://doi.org/10.1111/tpj.14904>
- 281 Hwang OJ, Back K (2021) Suppression of rice cryptochrome 1b decreases both melatonin  
 282 and expression of brassino steroid biosynthetic genes resulting in salt tolerance. *Molecules*  
 283 26(4):1075. [https://doi.org/10.3390/molecules26\(4\):1075](https://doi.org/10.3390/molecules26(4):1075)
- 284 Humann JL, Jung S, Cheng CH et al (2019) Cool season food legume genome database: a resource  
 285 for pea, lentil, faba bean and chickpea genetics, genomics and breeding. In: Proceedings of the  
 286 international plant and animal genome conference, January 2019, San Diego
- 287 Hyunmi K, Hyunsik H, Jung-WH et al (2012) A rice orthologue of the ABA receptor,  
 288 OsPYL/RCAR5, is a positive regulator of the ABA signal transduction pathway in seed germination  
 289 and early seedling growth. *J Exp Bot* 63(2):1013–1024. <https://doi.org/10.1093/jxb/err338>
- 290
- 291 J'afar U, Aliyu A, Kasimu S et al (2018) genetic diversity in tomato genotypes (*Solanum lycoper-*  
 292 *sicum*) based on salinity responsive candidate gene using simple sequence repeats. *Int Lett Nat*  
 293 *Sci* 72:37–46
- 294 Kong W, Zhong H, Deng X et al (2019) Evolutionary analysis of *GH3* genes  
 295 in six *Oryza* species/subspecies and their expression under salinity stress in *Oryza*  
 296 *sativa* ssp. *japonica*. *Plants* 8(2):30. <https://doi.org/10.3390/plants8020030>
- 297 Kumar P, Sharma PK (2020) Soil salinity and food security in India. *Front Sustain Food Syst* 4:174.  
 298 <https://doi.org/10.3389/fsufs.2020.533781>
- 299 Kurata N, Yamazaki Y (2006) Oryzabase. An integrated biological and genome information database  
 300 for rice. *Plant Physiol* 140(1):12–7. <https://doi.org/10.1104/pp.105.063008>
- 301 Lawrence CJ, Harper LC, Schaeffer ML (2008) Maize GDB: the maize model organism database  
 302 for basic, translational, and applied research. *Int J Plant Genomics*. <https://doi.org/10.1155/2008/496957>
- 303
- 304 Liu Z, Ge X, Yang Z et al (2017) Genome-wide identification and characterization of SnRK2 gene  
 305 family in cotton (*Gossypium hirsutum* L.). *BMC Genet* 18:54. <https://doi.org/10.1186/s12863-017-0517-3>
- 306
- 307 Liangbo F, Qiufang S, Lihui K (2018) Metabolite profiling and gene expression of Na/K transporter  
 308 analyses reveal mechanisms of the difference in salt tolerance between barley and rice. *Plant*  
 309 *Physiol Biochem* 130:248–257. <https://doi.org/10.1016/j.plaphy.2018.07.013>
- 310 Lizumi T, Sakai T (2020) The global dataset of historical yields for major crops 1981–2016. *Sci*  
 311 *Data* 7:97. <https://doi.org/10.1038/s41597-020-0433-7>

- 312 Macovei A, Sahoo RK, Faè M et al (2017) Over expression of PDH45 or SUV3 helicases in rice  
313 leads to delayed leaf senescence-associated events. *Protoplasma* 254(2):1103–1113. <https://doi.org/10.1007/s00709-016-1017-4>
- 314 Mantri NL, Ford R, Coram TE et al (2007) Transcriptional profiling of chickpea genes differentially  
315 regulated in response to high-salinity, cold and drought. *BMC Genomics* 8:303. <https://doi.org/10.1186/1471-2164-8-303>
- 316 Meng X, Li L, De Clercq (2019) ANAC017 Coordinates organellar functions and stress responses  
317 by reprogramming retrograde signalling. *Plant Physiol* 180(1):634–653. <https://doi.org/10.1104/pp.18.01603>
- 318 Moco S, Bino RJ, Vorst O (2006) a liquid chromatography-mass spectrometry-based metabolome  
319 database for tomato1. *Plant Physiol* 141:1205–1218
- 320 Muneer S, Jeong BR (2015) Proteomic analysis of salt-stress responsive proteins in roots of tomato  
321 (*Lycopersicon esculentum* L.) plants towards silicon efficiency. *Plant Growth Regul* 77:133–146
- 322 Najafi S, Sorkheh K, Nasernakhaei F (2018) Characterization of the APETALA2/Ethylene-  
323 responsive factor (AP2/ERF) transcription factor family in sunflower. *Sci Rep* 8(1):11576. <https://doi.org/10.1038/s41598-018-29526-z>
- 324 Narise T, Sakurai N, Obayashi T et al (2017) Co-expressed pathways database for tomato: a database  
325 to predict pathways relevant to a query gene. *BMC Genom* 18:437
- 326 NCBI Resource Coordinators (2016) Database resources of the National Center for Biotechnology  
327 Information. *Nucleic Acids Res* 44(D1):D7-19. <https://doi.org/10.1093/nar/gkv1290>
- 328 Nudrat AA, Fahad S, Muhammad A (2017) Ascorbic acid-a potential oxidant scavenger and its role  
329 in plant development and abiotic stress tolerance. *Front Plant Sci* 8:613. <https://doi.org/10.3389/fpls.2017.00613>
- 330 Oda Y, Kobayashi NI, Tanoi K (2018) T-DNA tagging-based gain-of-function of oshkt1;4 reinforces  
331 an exclusion from leaves and stems but triggers na toxicity in roots of rice under salt stress. *Int J Mol Sci* 19(1):235. <https://doi.org/10.3390/ijms19010235>
- 332 Pan YX, Yuan CF, Jing SY (2020) Simulation and optimization of irrigation schedule for summer  
333 maize based on SWAP model in saline region. *Int J Agric Biol Eng* 13(3):117–122
- 334 Platten, JD, Egdane, JA, Ismail, AM (2013) Salinity tolerance, Na<sup>+</sup> exclusion and allele mining  
335 of *HKT1;5* in *Oryza sativa* and *O. glaberrima*: many sources, many genes, one mechanism?. *BMC Plant Biol* 13:32. <https://doi.org/10.1186/1471-2229-13-32>
- 336 Reddy INBL, Kim BK, Yoon IS et al (2017) Salt tolerance in rice: focus on mechanisms and  
337 approaches. *Rice Sci* 24(3):123–144. <https://doi.org/10.1016/j.rsci.2016.09.004>
- 338 Rice Full-Length cDNA Consortium (2003) Collection, mapping, and annotation of over 28,000  
339 cDNA clones from japonica rice. *Science* 301(5631):376–379. <https://doi.org/10.1126/science.1081288>
- 340 Sampsel LJ (2018) Voyant tools. *Music Ref Serv Q* 21(3):153–157. <https://doi.org/10.1080/10588167.2018.1496754>
- 341 Sanderson LA, Caron CT, Tan R et al (2019) KnowPulse: a web-resource focused on diversity data  
342 for pulse crop improvement. *Front Plant Sci* 10:965. <https://doi.org/10.3389/fpls.2019.00965>
- 343 Sandhu M, Sureshkumar V, Prakash C et al (2017) RiceMetaSys for salt and drought stress responsive  
344 genes in rice: a web interface for crop improvement. *BMC Bioinform* 18:432. <https://doi.org/10.1186/s12859-017-1846-y>
- 345 Sato Y, Takehisa H, Kamatsuki K et al (2013) RiceXPro version 3.0: expanding the informatics  
346 resource for rice transcriptome. *Nucleic Acids Res* 41(Database issue):D1206–13. <https://doi.org/10.1093/nar/gks1125>
- 347 Shinozaki Y, Nicolas P, Fernandez-Pozo N et al (2018) High-resolution spatiotemporal transcriptome  
348 mapping of tomato fruit development and ripening. *Nat Commun* 9:364
- 349 Shirasawa K, Hirakawa H (2013) DNA marker applications to molecular genetics and genomics in  
350 tomato. *Breed Sci* 63(1):21–30. <https://doi.org/10.1270/jsbbs>
- 351 Shohat H, Illouz-Eliaz N, Kanno Y et al (2020) The tomato DELLA protein PROCERA promotes  
352 abscisic acid responses in guard cells by upregulating an abscisic acid transporter. *Plant Physiol*  
353 1184(1):518–528. <https://doi.org/10.1104/pp.20.00485>
- 354

- 365 Simon NML, Kusakina J, Fernandez-Lopez A et al (2018) The energy-signaling hub SnRK1 is  
 366 important for sucrose-induced hypocotyl elongation. *Plant Physiol* 176:1299–1310
- 367 Suresh BV, Roy R, Sahu K et al (2014) Tomato genomic resources database: an integrated repository  
 368 of useful tomato genomic information for basic and applied research. *PLoS ONE* 9:e86387
- 369 Tang Y, Qin S, Guo Y et al (2016) Genome-wide analysis of the AP2/ERF gene family in physic  
 370 nut and over expression of the *jcerf011* gene in rice increased its sensitivity to salinity stress. *Plos*  
 371 *One* 11(3): <https://doi.org/10.1371/journal.pone.0150879>
- 372 Tello-Ruiz MK, Naithani S, Gupta P et al (2021) Gramene 2021: harnessing the power of compar  
 373 ative genomics and pathways for plant research. *Nucleic Acids Res* 49(D1):1452–1463. [https://](https://doi.org/10.1093/nar/gkaa979)  
 374 [doi.org/10.1093/nar/gkaa979](https://doi.org/10.1093/nar/gkaa979)
- 375 Tranchida-Lombardo V, Aiese Cigliano R, Anzar I et al (2018) Whole-genome re-sequencing of  
 376 two Italian tomato landraces reveals sequence variations in genes associated with stress tolerance,  
 377 fruit quality and long shelf-life traits. *DNA Res* 25(2):149–160. [https://doi.org/10.1093/dnares/](https://doi.org/10.1093/dnares/dsx045)  
 378 [dsx045](https://doi.org/10.1093/dnares/dsx045)
- 379 Timothy AE, Erica GJ, Christopher GL et al (2007) BASC: an integrated bioinformatics system for  
 380 *Brassica* research. *Nucleic Acids Res* 35(11):870–873. <https://doi.org/10.1093/nar/gk1998>
- 381 To HTM, Nguyen HT, Dang NTM et al (2019) Unraveling the genetic elements involved in shoot  
 382 and root growth regulation by jasmonate in rice using a genome-wide association study. *Rice*  
 383 12:69. <https://doi.org/10.1186/s12284-019-0327-5>
- 384 Toda Y, Tanaka M, Ogawa D et al (2013) RICE SALT SENSITIVE3 forms a ternary complex with  
 385 JAZ and class-C bHLH factors and regulates jasmonate-induced gene expression and root cell  
 386 elongation. *Plant Cell* 25(5):1709–1725. <https://doi.org/10.1105/tpc.113.112052>
- 387 Kudo T, Terashima S, Takaki Y et al (2017) Plant express: a database Integrating OryzaEx  
 388 press and ArthaExpress for single-species and cross-species gene expression network analyses  
 389 with microarray-based transcriptome data. *Plant Cell Physiol* 58:1. [https://doi.org/10.1093/pcp/](https://doi.org/10.1093/pcp/pcw208)  
 390 [pcw208](https://doi.org/10.1093/pcp/pcw208)
- 391 Wang, Z, Tang, J, Hu, R et al (2015) Genome-wide analysis of the R2R3-MYB transcription  
 392 factor genes in chinese cabbage (*Brassica rapa* ssp. *pekinensis*) reveals their stress and hormone  
 393 responsive patterns. *BMC Genom* 16:17. <https://doi.org/10.1186/s12864-015-1216-y>
- 394 Wang H, Lei Y, Wan L et al (2016) Comparative transcript profiling of resistant and susceptible  
 395 peanut post-harvest seeds in response to aflatoxin production by *Aspergillus flavus*. *BMC Plant*  
 396 *Biol* 16:54. <https://doi.org/10.1186/s12870-016-0738-z>
- 397 Wang D, Cao Z, Wang W et al (2020) Genome-wide characterization of *OFP* family genes in  
 398 wheat (*Triticum aestivum* L.) reveals that *TaOPF29a-A* promotes drought tolerance. *Biomed Res*  
 399 *Int*7:9708324. <https://doi.org/10.1155/2020/9708324>
- 400 Wong LH, Gatta AT, Levine TP (2019) Lipid transfer proteins: the lipid commute via shuttles,  
 401 bridges and tubes. *Nat Rev Mol Cell Biol* 20:85–101. <https://doi.org/10.1038/s41580-018-0071-5>
- 402 Xie X, Zhang Z, Zhao Z et al (2020) The mitochondrial aldehyde dehydrogenase OsALDH2b  
 403 negatively regulates tapetum degeneration in rice. *J Exp Bot* 71(9):2551–2560. [https://doi.org/](https://doi.org/10.1093/jxb/eraa045)  
 404 [10.1093/jxb/eraa045](https://doi.org/10.1093/jxb/eraa045)
- 405 Xu H, Li X, Zhang H et al (2020) High temperature inhibits the accumulation of storage materials  
 406 by inducing alternative splicing of OsbZIP58 during filling stage in rice. *Plant Cell Environ*  
 407 43(8):1879–1896. <https://doi.org/10.1111/pce.13779>
- 408 Young UT, Yong LH, Sangyool L et al (2018) jasmonate zim-domain protein 9 interacts with slender  
 409 rice 1 to mediate the antagonistic interaction between jasmonic and gibberellic acid signals in  
 410 rice. *Front Plant Sci* 9:1866. <https://doi.org/10.3389/fpls.2018.01866>
- 411 Zhang H, Xiang Y, He N et al (2020) Enhanced vitamin C production mediated by an ABA-  
 412 Induced PTP-like nucleotidase improves plant drought tolerance in arabidopsis and maize. *Mol*  
 413 *Plant* 13(5):760–776. <https://doi.org/10.1016/j.molp.2020.02.005>
- 414 Zheng M, Lin J, Liu X et al (2021) Histone acetyltransferase TaHAG1 acts as a crucial regulator to  
 415 strengthen salt tolerance of hexaploid wheat. *Plant Physiol* 23:kiab187. [https://doi.org/10.1093/](https://doi.org/10.1093/plphys/kiab187)  
 416 [plphys/kiab187](https://doi.org/10.1093/plphys/kiab187)

417 Zhou S, Sauvé R, Thannhauser TW (2009) Proteome changes induced by aluminium stress in  
418 tomato roots. *Plant Signal Behav* 60:1849–1857

UNCORRECTED PROOF



# Author Queries

Chapter 5

Query Refs.	Details Required	Author's response
AQ1	Kindly note that the reference citation Xu et al. (2021) has been changed to Xu et al. (2020) so that this citation matches the Reference List. Please confirm that this is correct.	

UNCORRECTED PROOF

# MARKED PROOF

## Please correct and return this set

Please use the proof correction marks shown below for all alterations and corrections. If you wish to return your proof by fax you should ensure that all amendments are written clearly in dark ink and are made well within the page margins.

<i>Instruction to printer</i>	<i>Textual mark</i>	<i>Marginal mark</i>
Leave unchanged	... under matter to remain	Ⓟ
Insert in text the matter indicated in the margin	∧	New matter followed by ∧ or ∧ <sup>Ⓢ</sup>
Delete	/ through single character, rule or underline or ┌───┐ through all characters to be deleted	Ⓞ or Ⓞ <sup>Ⓢ</sup>
Substitute character or substitute part of one or more word(s)	/ through letter or ┌───┐ through characters	new character / or new characters /
Change to italics	— under matter to be changed	↵
Change to capitals	≡ under matter to be changed	≡
Change to small capitals	≡ under matter to be changed	≡
Change to bold type	~ under matter to be changed	~
Change to bold italic	≈ under matter to be changed	≈
Change to lower case	Encircle matter to be changed	≡
Change italic to upright type	(As above)	⊕
Change bold to non-bold type	(As above)	⊖
Insert 'superior' character	/ through character or ∧ where required	Υ or Υ under character e.g. Υ or Υ
Insert 'inferior' character	(As above)	∧ over character e.g. ∧
Insert full stop	(As above)	⊙
Insert comma	(As above)	,
Insert single quotation marks	(As above)	ʹ or ʸ and/or ʹ or ʸ
Insert double quotation marks	(As above)	“ or ” and/or “ or ”
Insert hyphen	(As above)	⊞
Start new paragraph	┌	┌
No new paragraph	┐	┐
Transpose	└┐	└┐
Close up	linking ○ characters	Ⓞ
Insert or substitute space between characters or words	/ through character or ∧ where required	Υ
Reduce space between characters or words		↑